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A FUROSTANOL SAPONIN FROM FRUITS OF BALANITES AEGYPTIACA

M. S. KAMEL*

Department of Pharmacognosy, Faculty of Pharmacy, Assiut University, Assiut, Egypt

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Key Word Index—Balanites aegyptiaca; Balanitaceae; balanitesin; furostanol saponin.

Abstract—A new furostanol saponin was isolated from the mesocarp of *Balanites aegyptiaca* fruits and identified as 26-O- β -D-glucopyranosyl-(25R)-furost-5-ene-3,22,26-triol 3-O- $\{[\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)]$ - β -D-xylopyranosyl- $(1 \rightarrow 2)\}$ - β -D-xylopyranoside (balanitesin). © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Balanites aegyptiaca Del. is a widely distributed African plant of Medicinal interest [1–4]. In Egyptian folk-medicine, the fruits are used as an oral hypoglycemic drug. Our previous work on this plant resulted in isolation and characterization of new hypoglycemic furostanol saponins in addition to new steroidal glycosides [5–7]. This paper deals with isolation and elucidation of a new furostanol saponin from the mesocarp of the fruits.

RESULTS AND DISCUSSION

The methanolic extract of the mesocarp of *Balanites* aegyptiaca fruits was defatted with *n*-hexane and fractionated on a column of Diaion HP-20 to yield com-

$$\begin{array}{c|c} xyl & O \\ \hline \\ Rha & 4 \\ 2 \\ 2 \\ 2 \\ Rha & Xyl \\ \end{array}$$

pound 1. Compound 1 was assumed to be a furostanol saponin on the basis of the positive coloration with Ehrlich reagent [8]. The acid hydrolysis of 1 vielded D-glucose, L-rhamnose and D-xylose in addition to diosgenin as the aglycone. The sugars were identified using TLC by comparison with authentic samples and also by GLC of their trimethylsilyl derivatives. A peak at m/z 1283 [M+H]⁺ in the positive ion FAB-mass was consistent with the molecular formula $C_{60}H_{98}O_{29}$. The following significant fragment peaks were also observed at m/z 1137 $[M+H-Rha]^+$, m/z 975 $[M+H-Rha-Glc]^+$, m/z 829 [M+H-2Rha-Glc]⁺ and m/z 433 [M+H- 2Rha-Glc-3Xyl]⁺. The carbon signals due to the aglycone moiety in the ¹³C NMR spectrum of 1 (Table 1) appeared at almost the same positions as those reported for the previously isolated 22-hydroxylated furostanol saponins from Balanites aegyptiaca [5, 6], indicating that compound 1 was also a 3,26-O-bisdesmosidic furostanol saponin with the same aglycone moiety. The carbon signals at δ 106.9, 104.6, 104.6, 104.4, 101.7 and 100.0 (Table I) were clear for the anomeric carbons of six sugar units which were identified as one terminal β -glucopyranosyl, one terminal α-rhamnopyranosyl, one terminal β -xylopyranosyl, one substituted α -rhamnopyranosyl and two substituted β -xylopyranosyl units respectively [9]. The β -configuration of the Dglucopyranosyl and D-xylopyranosyl units was indicated from the coupling constants (6.5 Hz, 7.0 Hz, 7.2 Hz and 7.0 Hz) of the doublet signals at δ 4.81, 5.10, 5.13 and 5.22 respectively in the ¹H NMR spectrum [10], while the α -configuration of the L-rhamnopyranosyl units was deduced from the downfield shifts of C-5 (δ 69.4 each) in the ¹³C NMR spectrum [9, 11]. The attachment sites of the substituted sugars have been determined by inspection of the other sugar signals which revealed the downfield shifts of C-2 of

^{*} Author to whom correspondence should be addressed.

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Table 1. ¹³C NMR spectral data of compound 1 (100 MHz, pyridine-d_s)

C	1	C	1	C	1
1	37.5*	* 22	110.7	Subst. Rha	
2	30.1	23	37.1	1	100.0
3	78.4	24	29.0	2	78.8
4	38.9	25	35.1	3	71.2
5	140.8	26	75.2	4	73.9
6	121.8	27	17.4	5	69.4
7	32.3	26-Glc		6	18.6
8	31.7	Ī	104.4	Term. Xyl	
9	50.1	2	75.3	1	106.9
10	37.1*	* 3	78.6	2	75.4
11	21.1	4	71.7	3	78.0
12	40.01	÷ 5	78.2	4	70.9
13	40.3†	· 6	62.8	5	68.5
14	57.0	Term. Rha		Subst. Xyl	
15	32.8	1	101.7	1,1'	104.6, 104.6
16	81.2	2	72.4	2,2'	81.6, 81.6
17	62.1	3	72.7	3,3′	77.3, 76.1
18	16.4	4	74.9	4,4′	70.9, 78.0
19	19.3	5	69.4	5,5'	68.5, 67.3
20	40.7	6	18.6		
21	16.4				

^{*†} Values may be interchangeable in each column.

the substituted rhamnopyranosyl unit and one of the substituted xylopyranosyl units (δ 78.8, 81.6 respectively) in addition to the downfield shifts of C-2 and C-4 (δ 81.6 and 78.0, respectively) of the second substituted xylopyranosyl unit [11]. The full assignment of the sugar moiety was established by 'H detected Multiple Bond Connectivity (HMBC) based on the assignment of the sugar proton and carbon signals by ¹³C-¹H Correlation Spectroscopy (2D COSY). The correlations (Fig. 1) were obvious between H-1 of the 2,4-disubstituted xylopyranosyl unit and C-2 of the 2substituted xylopyranosyl unit, H-1 of the terminal xylopyranosyl and C-2 of the 2,4-disubstituted xylopyranosyl unit, H-1 of the 2-substituted rhamnopyranosyl unit and C-4 of the 2,4-disubstituted xylopyranosyl unit in addition to H-1 of the terminal rhamnopyranosyl unit and C-2 of the 2-substituted rhamnopyranosyl unit. Consequently, compound 1 was assigned the structure 26-O- β -D-glucopyranosyl-(25R)-furost-5-ene-3,22,26-triol 3-O- $\{[\alpha$ -L-rham-nopyranosyl- $(1 \rightarrow 2)$ - α L-rham-nopyranosyl- $(1 \rightarrow 4)$]- $[\beta$ -D-xylopyranosyl- $(1 \rightarrow 2)$]- β -D-xylopyranosyl- $(1 \rightarrow 2)$ }- β -D-xylopyranoside (balanitesin).

EXPERIMENTAL

NMR spectra were recorded in pyridine-d₅ using a JEOL JNM-GX-400 spectrometer (400 MHz for ¹H NMR and 100 MHz for ¹³C NMR) with tetramethylsilane (TMS) as internal standard. The FABmass was recorded by a Kratos MS 80RF mass spectrometer. GLC conditions: Perkin-Elmer GC apparatus equipped with flame ionization detector, carrier gas He (3 ml min⁻¹), Neutra Bond-1 column (0.25 mm i.d. × 30 m), column temp. 180°, injection temp. 225°. R_t L-rhamnosc 6.0 min, D-xylose 7.9 min and Dglucose 21 min. TLC was carried out on precoated silica gel plates (Kieselgel 60 F₂₅₄, Merck). For CC, silica gel G (E. Merck), Lichroprep RP-8 (40-63 μ m, Merck) and Diaion HP-20 (Mitsubishi Chem. Ind. Co., Ltd) were used. The solvent systems were: (I) CHCl₃-MeOH-H₂O (65:35:10, lower phase); (II) CHCl₃-MeOH-H₂O (60:40:10); (III) 70% MeOH and (IV) 65% MeOH. Spray reagents: 10% H₂SO₄ and triphenyltetrazolium chloride (TTC). Plant material was collected in May 1993 from Al-Wady Al-Gadid area, western desert of Egypt. The identity of the plant was confirmed by Prof. A. Fayed, Faculty of Science, Assiut University. A voucher sample is kept in the Herbarium of the Faculty of Pharmacy, Assiut University, Egypt.

Extraction and isolation

The mesocarp (750 g) of *B. aegyptiaca* fruits was extracted with MeOH. After removal of the solvent by evapn the residue was defatted with *n*-hexane. The mother liquor was chromatographed on Diaion HP-20 and eluted with H₂O, 40% MeOH, 80% MeOH, MeOH and Me₂CO successively. The 40% MeOH eluate was chromatographed on silica gel (system II) affording three fractions. Fraction 2 was subjected to CC on RP-8 using system III followed by prep. HPLC

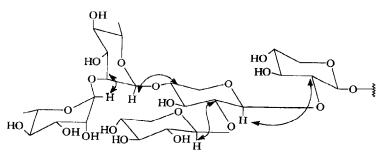


Fig. 1. HMBC Correlations of 1.

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on an ODS column (system IV) to provide compound 1.

Compound 1 (balanitesin). Powder (50 mg). $[\alpha]_d^{23} + 25.2^{\circ}$ (MeOH; c 0.31) and R_f 0.12 (system I). ¹H NMR (pyridine- d_s): δ 0.80 (3H, s, Me-18), 0.88 (3H, d, J = 7.0 Hz, Me-27), 1.02 (3H, s, Me-19), 1.4 (3H, d, J = 6.8 Hz, Me-21), 1.75 (6H, d, J = 6.1 Hz, 2Me-6 Rha), 4.81 (1H, d, J = 6.5 Hz, H-1 Glc), 5.10 (1H, d, J = 7.0 Hz, H-1 Xyl), 5.13 (1H, d, J = 7.2 Hz, H-1 Xyl), 5.22 (1H, d, J = 7.0 Hz, H-1 Xyl), 6.22 (1H, d, d = 7.0 Hz, H-1 Rha) and 6.24 (1H, d = 7.1 Rha). FAB-mass (positive) $m_f z$: 1283 [C₆₀H₉₈O₂₉ + H]⁺, 1137, 975, 829 and 433.

Acid hydrolysis

A sample (30 mg) was heated with 2 M HCl in H₂O-dioxane (1:1) in a sealed tube at 80° for 3 h. The reaction mixture was diluted with H₂O and extracted with CHCl₃. The aq. layer was neutralized with Ag₂CO₃ and subjected to TLC on silica gel using system II (detection: TTC reagent). *R_f* values were 0.17, 0.32 and 0.25 for D-glucose, L-rhamnose and D-xylose, respectively. The aq. layer was concd (20 mg), trimethylsilylated and subjected to GLC analysis. The identification of the aglycone (diosgenin) isolated from the CHCl₃ layer was based on ¹³C NMR analysis and comparison with reported data [9].

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